



## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<b>(51) International Patent Classification 5 :</b>  <b>A61L 27/00</b>	<b>A1</b>	<b>(11) International Publication Number:</b> <b>WO 93/00935</b>  <b>(43) International Publication Date:</b> 21 January 1993 (21.01.93)
<b>(21) International Application Number:</b> PCT/GB92/01247 <b>(22) International Filing Date:</b> 9 July 1992 (09.07.92)  <b>(30) Priority data:</b> 91306305.3 11 July 1991 (11.07.91) EP <b>(34) Countries for which the regional or international application was filed:</b> AT et al.  <b>(71) Applicant (for all designated States except US):</b> BRITISH TECHNOLOGY GROUP LTD. [GB/GB]; 101 Newington Causeway, London SE1 6BU (GB).  <b>(72) Inventors; and</b> <b>(75) Inventors/Applicants (for US only) :</b> BEERSTEN, Wouter [NL/NL]; Anna Van Burenlaan 21, NL-2082 BK Santpoort (NL). VAN DEN BOS, Theo [NL/NL]; Brugakker 1309, NL-3704 WE Zeist (NL).		<b>(74) Agent:</b> HAMILTON, Raymond; Patents Department, British Technology Group Ltd., 101 Newington Causeway, London SE1 6BU (GB).  <b>(81) Designated States:</b> JP, US, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LU, MC, NL, SE).  <b>Published</b> <i>With international search report.</i>
<b>(54) Title:</b> IMPLANT MATERIALS  <b>(57) Abstract</b>  <p>The combination of a phosphatase enzyme with a biocompatible carrier material produces materials which are useful in the repair of the skeleton and the promotion of new bone growth. The combination preferably involves covalent coupling between the enzyme and the carrier. The preferred carrier materials comprise fibrillar collagen and may be obtained by the demineralisation of calcified tissues. The materials may comprise phosphoproteins or dentinal phosphophoryns which may be residual or may be added during the preparation of the materials. The incorporation of other organophosphates and inorganic phosphates may improve the rate of mineralisation especially in older animals.</p>		

**FOR THE PURPOSES OF INFORMATION ONLY**

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	FI	Finland	ML	Mali
AU	Australia	FR	France	MN	Mongolia
BB	Barbados	GA	Gabon	MR	Mauritania
BE	Belgium	GB	United Kingdom	MW	Malawi
BF	Burkina Faso	GN	Guinea	NL	Netherlands
BG	Bulgaria	GR	Greece	NO	Norway
BJ	Benin	HU	Hungary	PL	Poland
BR	Brazil	IE	Ireland	RO	Romania
CA	Canada	IT	Italy	RU	Russian Federation
CF	Central African Republic	JP	Japan	SD	Sudan
CG	Congo	KP	Democratic People's Republic of Korea	SE	Sweden
CH	Switzerland	KR	Republic of Korea	SN	Senegal
CI	Côte d'Ivoire	LJ	Liechtenstein	SU	Soviet Union
CM	Cameroon	LK	Sri Lanka	TD	Chad
CS	Czechoslovakia	LU	Luxembourg	TG	Togo
DE	Germany	MC	Monaco	US	United States of America
DK	Denmark	MG	Madagascar		
ES	Spain				

- 1 -

IMPLANT MATERIALS

This invention relates to novel materials useful in the restoration and repair of the skeletal system, to processes for their production and to novel methods for the repair of the skeleton which utilise them.

There are a variety of surgical procedures used in man and animals to repair the skeleton. These include the healing of fractures by insertion of an implant to stabilise and facilitate healing of the fracture, the implantation of bone substitute materials, especially in those areas of the body which show a low intrinsic tendency to heal and the insertion of an implant which serves as a membrane to regenerate supportive tissues. The materials used as implants are ideally biocompatible and of a nature such as will promote the growth of bone.

The possible role of the enzyme alkaline phosphatase in promoting the calcification of bone has been postulated for many years. Recent studies notably an in vitro study reported by Beertsen and Van Den Bos (Matrix Vol. 9 1989 p159-171) have tended to support the proposition that alkaline phosphatase may be involved in the initiation of the calcification process apparently by virtue of its acting so as to raise the local concentration of phosphate ions.

However, the relevance of such in vitro mineralisation studies to the situation in vivo has been questioned, particularly in view of the relatively high concentrations of phosphate esters used in the in vitro studies and also because the rate of hydrolysis of the phosphate esters at physiological pH levels would be expected to be too low to be relevant to the process of mineralisation.

USP 4,394,370 describes a bone graft material which comprises a complex of reconstituted collagen and either demineralised bone particles or a solubilised bone morphogenetic protein fabricated as a sponge. These sponges may be complexed with bovine intestinal alkaline phosphatase. A concentration of

- 2 -

15mg per gram of collagen dispersion is said to eliminate all inflammatory responses to the graft material, accelerate the formation of osteoid in the graft material and aid the slow resorption of the graft enabling it to be more completely  
5 corticalised. There is no mention of any mineralisation having occurred. USP 4,409,332 describes porous membrane structures based on collageneous materials which are complexed with alkaline phosphatase in order to reduce the inflammatory reaction produced when these membranes are introduced into the  
10 body.

We have now discovered that novel materials useful in the repair of the skeleton may be produced by combining a biocompatible carrier material with a quantity of a phosphatase enzyme such as will promote mineralisation. The level of enzyme  
15 activity is preferably at least 0.5 milliunits Apase per 1.0 $\mu$ gm of hydroxyproline (where one unit is defined as 1 $\mu$ mol p-nitrophenol released per minute from p-nitrophenyl phosphate at 37°C and pH 10.5 using the technique described by Beertsen and van den Bos in Matrix, Vol. 9/1989, p161).

20 Biocompatible carrier materials may themselves mineralise at least to some degree if used to repair the skeleton and allowed to remain in situ for an extended period. The materials of this invention promote mineralisation of the implant so that it can be readily detected within seven days and have practical utility  
25 in the repair of the skeleton.

The novel materials are stable products which when used to repair the skeleton undergo mineralisation and may serve to promote the growth of new bone. Accordingly, from one aspect this invention provides a novel material useful in the repair of  
30 the skeleton which comprises a phosphatase enzyme combined with a bio-compatible carrier material and having a level of enzyme activity as will promote mineralisation.

- 3 -

The phosphatase enzyme is preferably an alkaline phosphatase enzyme (hereinafter for convenience "APase"). APase is normally obtained by extraction from human or animal tissue. The enzyme is a cell surface glycoprotein which is capable of hydrolysing a variety of monophosphate esters. Three main forms of APase are distinguished; liver/bone/kidney; placental and intestinal and all of these forms are useful in this invention.

The carrier material may comprise any biocompatible material which is capable of combining with the enzyme. Preferably, the combination is brought about by incubating the carrier with the enzyme in the presence of a coupling agent which is capable of covalently bonding with the carrier and with the enzyme. Suitable coupling agents are those which are capable of bonding to the enzyme without significantly reducing its biological activity. A wide variety of polyfunctional compounds may be useful. The coupling agents used to form the preferred materials of this invention will be those which are capable of bonding to the carrier material. Examples of potentially useful coupling agents include biotin-avidin; glutaraldehyde and 1-ethyl-3(3-dimethyl-aminopropyl) carbodiimide HCl. The preferred coupling agents are those which do not produce an adverse reaction when introduced into the body in particular as part of a material according to this invention. The most preferred coupling agents are the carbo-diimides and in particular the compound mentioned above.

A particularly preferred coupling agent is that known as SATA-MHS, which involves the use of a combination of succinimidyl-S-acetylthioacetate (SATA) and maleimido hexanoyl-N-hydroxysuccinimide (MHS). Preferably the carrier is incubated with the SATA and the enzyme with the MHS. The products of these two incubation processes are combined and allowed to react to produce an implant material.

- 4 -

The carrier material may be of natural or synthetic origin. A wide variety of synthetic polymers are potentially useful. A particular group of synthetic materials which may be of use are those polymers which are known to be bioabsorbable. The use of  
5 synthetic materials is also advantageous insofar as they may be flexible and capable of being formed into shaped pieces designed for particular applications.

A variety of natural materials may also be used as the carrier. In particular human or animal tissues such as bones  
10 and teeth may be useful. Calcified tissues such as bones and teeth must be demineralised before they can be used as a carrier. The materials obtained by demineralisation comprise a substantial proportion of fibrillar collagen. Collagen is the major fibrous protein of many animals and may be extracted from  
15 many parts of the human or animal body. Fibrillar collagen is a preferred carrier material for use in the present invention as are materials which comprise a substantial proportion of it. An example of such a material is human or animal dura mater. The carrier materials are preferably those which are sufficiently  
20 flexible to be formed into shaped pieces. Those materials which are porous or comprise meshes of fibrillar or fibrous materials are also advantageous in that they have a larger surface area to which the enzyme may be bonded. Useful carrier material may also be formed by incorporating collagen fibres into or onto a  
25 suitable supporting material always provided that the surface of the fibres remains exposed. An example of a supporting material is natural or synthetic calcium phosphates which substances may usefully form part of the materials of this invention.

The carrier material will preferably be sufficiently  
30 flexible and sufficiently strong to be useful as an implant material. The degree of flexibility or strength may vary according to the particular application for which the material is intended. The material is preferably one which is capable of being handled and manipulated prior to and during the

- 5 -

implantation operation. The preferred implant materials comprise at least 200 units of phosphatase per cubic centimetre. The preferred carrier materials are those which are capable of combining with this amount of enzyme. Useful  
5 materials may be produced by compression of a carrier or of a material produced by combination of a carrier and a phosphatase enzyme.

The preferred carrier materials are those having a relatively smooth surface and a densely organised fibrous  
10 structure. Such materials may exhibit osteoconductive properties, i.e. they appear to guide the migration of osteoblasts and thereby to encourage the growth of new bone.

It is also preferred that the carrier has a tubular structure, i.e. one having a series of cracks or  
15 microfractures. The enzyme is bound within these tubules and mineralisation may occur in them. This is advantageous insofar as the strength of the material increases as the mineralisation proceeds. Mineralisation may be enhanced by pre-incubating the carrier (or the combined carrier - APase material) in a solution  
20 containing physiological concentrations of calcium and an organophosphate (e.g. a  $\beta$ -glycerophosphate solution) or an inorganic phosphate. The pre-incubation appears to form nucleation centres within the carrier which after implantation in the body may grow rapidly and boost the rate of mineralisation.

25 The carrier material is preferably one which is free from any substance which might serve to inhibit its mineralisation. In the case of materials of natural origin it may be necessary to extract the material thoroughly in order to reduce the level of any inhibitor to an acceptable value. Conveniently  
30 demineralisation of a calcified tissue may be effected by washing the cleaned tissue and placing it in an acid solution, e.g. of acetic or hydrochloric acid for an extended period. Preferably the resulting demineralised material is further extracted with a chaotropic agent in the presence of a chelating  
35 agent.

- 6 -

The demineralised material may contain residual quantities of non-collagenous proteins and insoluble phosphoproteins or dentinal phosphophoryns. The presence of bound phosphoproteins or dentinal phosphophoryns has been discovered to be efficacious in promoting the remineralisation. The preferred carrier materials are those which inherently comprise such phosphoproteins or phosphophoryns or those into which such phosphoproteins or phosphophoryns have been introduced and preferably covalently bound to the carrier. Preferably the materials of this invention comprise at least 0.03 micrograms of phosphate per microgram of hydroxyproline most preferably in the form of phosphoproteins or phosphophoryns.

The novel materials of this invention may be produced by incubating the carrier with the APase in the presence of the coupling agent and optionally in the presence of an organo-phosphorus compound.

The coupling reaction may conveniently be carried out by introducing the carrier and the APase into a solution of the coupling agent. The reaction should be carried out under conditions which do not inactivate the APase. Conveniently allowing the reactants to stand at a temperature which is not greater than ambient and is preferably less than 10°C for periods which will in general be at least 1 hour and may conveniently be longer, say 24 hours, will be sufficient to produce a product according to the invention.

The amount of APase present in the novel materials may vary within wide limits. Preferably the materials particularly those provided that it is sufficient to promote remineralisation in the intended application will exhibit a level of enzyme activity which is at least 1.0 milliunit Apase per 1.0µgm of hydroxyproline. The optimum amount of enzyme to be incorporated will vary with the nature of the component materials, the use to which the material is to be put, the age of the person or animal into which it is to be implanted and the concentration of inorganic phosphate present in the serum of the person or animal.



- 7 -

Materials comprising lower amounts of enzyme may be useful in animals which are undergoing a period of relatively rapid growth. In mature animals the implants preferably contain a higher concentration of enzyme most preferably in conjunction  
5 with phosphoproteins and phosphophoryns.

Alternatively, it is possible to promote mineralisation by increasing the concentration of phosphate in the locality of the implants, e.g. by transcutaneous or intravenous administration of organophosphates or inorganic phosphates. Methods of  
10 treatment of the skeleton in order to repair it which comprise the introduction of an implant material as hereinbefore described form another aspect of this invention. Methods of treatment further comprise an increase of the level of phosphates in the locality of the implant from a preferred  
15 aspect.

The amount of coupling agent employed will simply be that required to bind the desired amount of APase. In general the coupling agent will be present in the solution in a large excess over the quantity required by stoichiometry. The presence of  
20 such excess quantities of coupling agent may act so as to cross-link the surface of the carrier. Such cross-linking may be useful in improving the mechanical properties of a carrier material. It may also be useful in regulating the biodegradability of the implant. The implant should retain its  
25 mechanical integrity during the mineralisation but may usefully degrade thereafter.

The materials of this invention find use in a variety of surgical procedures. The carrier material will preferably be formed into an appropriate shape prior to it being coupled to  
30 the APase.

In particular the material finds use as an internal wound dressing to stabilise and facilitate the healing of fractures or defects of the skeleton. Following implantation it mineralises and hardens. The novel materials may also be used as a  
35 substitute for bone and used to repair fractures by fixing the

- 8 -

material into position and allowing it to mineralise and harden in situ. The treatment of fractures in these ways provides a further aspect of this invention.

The materials also find use in treatment processes which involve what has been termed "guided tissue regeneration". Such procedures involve the insertion of a membrane to help regenerate supportive connective tissues in areas of the body which have been damaged by disease. The membranes currently used in this application are often not biodegradable and must be removed in a second surgical operation. The materials of this invention are advantageous in this application insofar as they are biocompatible and may be allowed to remain in situ thus avoiding the need for further surgical intervention.

The invention is illustrated by the following examples.

15 Example 1

Preparation of dentinal collagen sheets. Bovine permanent incisors were collected at the local slaughterhouse immediately after killing (age 1-3 yr) and frozen at -80°C until use. After defrosting, the gingiva and periodontal ligament were removed and the roots cut with a diamond disk parallel to their longitudinal axis from the apex to the cervical area under constant irrigation with tap water and split with a chisel. The roots were then cleaned and freed from pulp and cementum. They were washed with ice-cold PBS in the presence of proteinase inhibitors and the outer dentin (containing the mantle dentin layer) was removed with a diamond disk under cooling with tap water.

Demineralised dentin slices (DDS) were prepared by soaking in 0.5M acetic acid or 0.6M hydrochloric acid for four weeks. Sections were cut by means of a cryotome set at 30µm. They were then further extracted with 4M guanidine.HCl and 0.4M EDTA (pH 7.5) for three days at 4°C. Before use the DDS were washed in double-distilled water for one hour and placed in double-distilled water supplemented with antibiotics at 4°C overnight. DDS contained 0.04µg PO<sub>4</sub> per µg hydroxyproline.

- 9 -

Binding of APase. Bovine intestinal APase was covalently bound to the dentinal collagen sheets by using the coupling agents glutaraldehyde or carbodiimide.

Glutaraldehyde coupling. The DDS were incubated in PBS  
5 containing 0.1% glutaraldehyde for 2h at 25°C in the presence of APase (700 U per ml). The material was then extensively washed with PBS and stored at 4°C in 0.1M glycine buffer pH 10.5 containing 1mM  $Mg^{2+}$  and 0.1mM  $Zn^{2+}$ .

Carbodiimide coupling. The DDS were incubated for 2 days at  
10 4°C in 0.13M 1-ethyl-3(3-dimethylaminopropyl)carbodiimide.HCl pH 4.5 in the presence of APase (700 U per ml). They were then exhaustively washed with distilled water, with 1M NaCl in 0.1M Na-acetate pH 4.0, with distilled water, with 0.1M  $NaHCO_3$  pH 8.3 and finally with distilled water. The material was stored in  
15 glycine buffer at 4°C (see preceding paragraph). The enzyme retained its activity under these storage conditions for at least 10 months.

In vitro experiments. In order to study the deposition of mineral in the collagen sheets as a function of time in vitro,  
20 APase-treated DDS (30 $\mu$ m thickness) and their controls (treated with crosslinking agents only or heat inactivated enzyme) were incubated in Iscove Modified Dulbecco's Medium (IMDM) supplemented with 10% heat-inactivated Normal Rabbit Serum (NRS) and antibiotics for varying time periods at 37°C. Radiolabelled  
25 calcium (1 $\mu$ Ci [ $^{45}Ca$ ]Cl<sub>2</sub> per well) was added and the collagen sheets monitored for uptake of the label. As phosphate source  $\beta$ -glycerophosphate ( $\beta$ -GP) was added in a concentration of 10mM. At the end of the experiment the specimens were decalcified in 0.5ml 1M HCl for 1h at 37°C. Samples of 300 $\mu$ l were added to  
30 Optifluor scintillation cocktail (Packard Instruments Inc.) and counted in a Packard Tricarb 4530 scintillation counter.

In vivo experiments. APase-treated DDS was tested for its ability to calcify under in vivo conditions as follows. Female Wistar rats (about 200g each) were anaesthetized with Hypnorm  
35 and an incision was made through the skin covering the skull

- 10 -

following the sagittal suture. The skin was retracted so as to expose the right and left temporal muscles. A pouch was then created on either side by cutting through the insertion of the exposed muscle. DDS slices of 30 $\mu$ m thickness and 1cm in width, 2cm in length were inserted into the pouches: APase-treated ones on the right side and controls on the left side. Care was taken that only a narrow rim of the implant was in contact with the muscle; its greater part (90%) was in direct contact with the dermis. The skin wound was closed with nylon sutures and allowed to heal for time periods varying from one to four weeks. The animals were then anaesthetized again, the wounded region re-opened and the implants excised. A small portion of each implant was removed together with the surrounding connective tissue and prepared for histological examination. The remaining portion was freed from surrounding tissues and used for chemical analyses.

### Results

In vitro experiments. DDS treated with APase accumulated radiolabelled calcium when incubated in the presence of  $\beta$ -GP. In the absence of the monophosphate ester very little [ $^{45}\text{Ca}$ ] was found in the dentin. When incorporation of the label in carbodiimide-treated specimens was followed as a function of time, a rapid influx of [ $^{45}\text{Ca}$ ] was observed during the first day. Thereafter, a more gradual increase was noted. Control sheets, treated with carbodiimide without enzyme, remained almost free of radioactivity. APase-containing slices hardened rapidly and stained positively with the Von Kossa method. The mineral was laid down in the form of needle-like crystallites in close association with the collagen fibrils of the carrier and (according to X-ray diffraction) exhibited hydroxyapatite-like characteristics.

In vivo experiments. Healing of the skin wounds was uneventful. Inspection of the sites of implantation during the re-entry procedure revealed that, particularly with the carbodiimide-treated implants, the surrounding tissues were rich

- 11 -

in collagen and not inflamed. The glutaraldehyde-treated implants, however, were often surrounded by a richly vascularized, somewhat oedematous connective tissue. In both implant types the (APase-coupled) DDS had become hard within one  
5 week and acquired an opaque appearance. Control specimens were soft and translucent upon macroscopic inspection.

Chemical analysis demonstrated that mineral uptake by APase-treated DDS was very rapid at the early stages after  
10 installment. As time progressed a further increase occurred with respect to the carbodiimide-treated specimens. According to linear regression analysis, this increase was statistically significant for phosphate ( $p < 0.05$ ). After 4 wk, in carbodiimide-coupled specimens the calcium content per  $\mu\text{g}$  hydroxyproline was about 80% of that found in normal bovine  
15 dentin, while the phosphate content was about 60%.

When expressed in terms of molar concentrations, it appeared that the Ca/P ratio in the remineralized dentin was about 2.00 for the carbodiimide-treated specimens and 1.80 for the glutaraldehyde-treated ones. In normal bovine dentin this ratio  
20 was 1.7

#### Example 2

A series of implant materials were prepared using the materials and techniques described in Example 1. The coupling agent was 1-ethyl-3(3-dimethylaminopropyl)carbodiimide.

25 Forty female Wistar rats (body weight, ca. 200g) were anesthetized with Hypnorm. An incision was made through the skin parallel to the sagittal suture. The skin covering the right and left parietal bones was reflected so as to expose the cranial vault from the frontal to the occipital region. A  
30 periosteal flap was raised on either side following incisions along the sagittal, frontal and occipital sutures. By use of a slowly rotating trephine bur ( $\emptyset$  1.8mm) mounted in a dental handpiece, two through and through defects were made in each of the parietal bones leaving a distance of about 1.0 to 1.5mm in  
35 between. To prevent overheating during surgery, the tissues

- 12 -

were kept moistened with physiological saline. Special care was taken not to injure physically the underlying dura mater.

The wounds were covered with a graft, one on either side of the skull, each measuring 0.5cm in width and 0.8cm in length. 5 On the right side an APase-containing one was implanted, on the left side a control one (no enzyme). The periosteum and skin were closed. The animals were killed by decapitation at the following time intervals post surgery: 3 weeks, 6 weeks, 9 weeks and 12 weeks. The calvariae were dissected out and immersed in 10 a solution of 4% paraformaldehyde and 1% glutaraldehyde in 0.1 mol/L Na-cacodylate buffer (pH 7.4) for 24h. The specimens were postfixed in 1% OsO<sub>4</sub> and embedded in epoxy resin.

Each skull half was oriented on the microtome so as to section the two wounds in the same plane, perpendicular to the 15 cranial vault. Sections were stained with methylene blue or according to the Von Kossa method. From each specimen one methylene blue stained section was taken from the central area of the two defects and used for histomorphometric analysis.

Ultrathin sections were cut with a diamond knife, stained 20 with uranyl acetate and lead citrate and examined in a Zeiss EM 10C electron microscope.

Histomorphometric analysis. Of each selected section a tracing was made by using a sign prism at a magnification of x173. The mineralised area of each graft was measured by using 25 an X-Y device and expressed as percentage of the total area occupied in the plane of sectioning. Also the thickness of the grafts was assessed and the length of direct contact with newly formed bone was measured, both along the inner (directed towards the skull) and outer (directed towards the skin) aspects of the 30 grafts.

### RESULTS

As shown histochemically, APase-activity in the enzyme-treated implants was bound to the wall of the dentinal tubules and the outer aspects of the sheets. The distribution of the 35 activity was uniform, in that along the entire length of the

- 13 -

5 sheets the same staining pattern was seen. Sections of control sheets that were incubated with the crosslinking agent in the absence of APase did not show any histochemical staining. Based on the conversion rate of p-nitrophenylphosphate, the enzyme-treated collagen sheets contained  $0.95 \pm 0.24$  mU of enzyme per  $\mu\text{g}$  hydroxyproline.

10 Healing of the surgical sites was uneventful in that the inflammatory response of the tissues surrounding the grafts, as assessed macroscopically, was negligible. More specifically, there were no signs of edema, redness and swelling. Macroscopic inspection and careful palpation with tweezers of the implants after fixation revealed that the APase-containing grafts had hardened, whereas the controls had remained soft.

15 Light and electron microscopic examination demonstrated that the experimental grafts had accumulated mineral during the course of the experiment. Within the first three weeks following implantation about 55% of their sectioned area was occupied by mineral. Thereafter, the amount of mineral showed a slight further increase to about 70-80%. Areas that had remained free of mineral were often near the edges where the grafts were in contact with the connective tissues of the overlying periosteum. No conspicuous differences in mineral density were observed between parts of the graft overlying the osseous defects and those overlying intact bone.

25 In the control grafts no substantial mineralisation could be detected up to and including the nine week time interval. Twelve weeks after surgery, however, some of the control specimens contained mineralised areas, more or less randomly distributed along the grafts. On the average these areas occupied about 20% of the grafted material.

30 In all cases the mineral was deposited as small crystals in close association with the collagenous fibrils constituting the grafts. The dentinal tubules had remained open and along the edges of the grafts no layers of mineralised material had been

- 14 -

deposited in the adjacent periosteum. Also no signs of cartilage formation were found.

As the grafts mineralised osteoblasts appeared which contained an extensive Golgi apparatus and rough endoplasmic reticulum. In between the osteoblast layer and the mineralised part of the graft osteoid and bone was deposited. Although no measurements were carried out on the strength of attachment between implant and bone, the orientation of cracks due to sectioning artifacts in the mineralised matrices gave some indication. They were seldom found along the borderline between the two. Usually they passed the bone-implant boundary.

All these phenomena were particularly apparent along the inner aspect of the enzyme-treated collagen sheets, facing the calvarial bone. Along the outer aspect directed towards the ectocranial periosteal layer very little, if any, bone was formed. Correlation analysis showed that, as mineralisation of the grafts increased, also the length of direct bone contact along its inner aspect tended to increase (correlation coefficient  $r = 0.53$ ,  $n = 37$ ;  $p < 0.005$ ).

Also in the control implants a positive correlation was found between their degree of mineralisation and the length of direct contact with newly formed bone ( $r = 0.45$ ,  $n = 37$ ;  $p < 0.01$ ). However, the extent to which the grafts were associated with bone was far less than on the experimental side of the skull. The difference among the two sides proved to be statistically significant ( $p < 0.01$ ).

Little inflammatory reactions were seen in the tissues surrounding the grafted material. More specifically, plasma cells were virtually absent and polymorphonuclear leukocytes and lymphocytes were very sparse. However, all implants showed evidence of a mild foreign body reaction, in that macrophages and multinucleated giant cells were present along the implant surfaces, except where the implants were in direct contact with newly formed bone. This was the case on the control as well as the experimental sides. The number of giant cells along the



- 15 -

grafts was approximately the same for the experimentals and controls (Wilcoxon-signed-ranks test,  $p > 0.05$ ) and they were not specifically associated with either mineralised or non-mineralised parts of the grafts.

5 Despite the presence of multinucleated giant cells in relation to the grafts, no major resorption of the implants had occurred, neither in the non-mineralised nor in the mineralised areas. Evidence for this was provided by the observation that the grafts kept their overall width during the course of the  
10 experiment ( $32\mu\text{m}$ ) and were not invaded to a great extent by resorbing cells. Yet, at various sites cells were seen with cytoplasmic processes penetrating deeply into the dentinal tubules. In none of the mineralised grafts osteoclasts, exhibiting ruffled borders were observed and no Howship's  
15 lacunae were seen.

#### Example 3

Mineralisation of an implant material was studied as a function of age, sex, site of implantation and phosphoprotein content.

20 Male and female Wistar rats, 5 or 20 weeks old, were anaesthetised and their weight determined. Sheets of material were then implanted subcutaneously in the following sites: over the skull (parietal and frontal bones), the lower back region, the wall of the abdomen. Control slices were without APase or  
25 with heat-inactivated APase. After two weeks the animals were weighed again, killed (under general anesthesia) by decapitation and the sheets taken out and analysed for phosphate content. Samples of serum were assayed for inorganic phosphate.

Preparation of material. Two types of material were tested:  
30 one made from demineralised and guanidine-extracted bovine dentin (which still contained some covalently bound dentinal phosphophoryns:  $0.035 \pm 0.019\mu\text{g phosphate}/\mu\text{g hypro}$ ) and the other made from demineralised and guanidine-extracted bovine cortical bone ( $0.005 \pm 0.0025\mu\text{g phosphate}/\mu\text{g hypro}$ ). Slices  
35 were made as described in Example 1.

- 16 -

APase was bound to the collagenous carrier (mainly type I collagen) according to the SATA-MHS coupling method.

A: SATA (succinimidyl-S-acetylthioacetate, Pierce) was linked to the carrier:

5        175mg of collagen was incubated in 5ml 0.05M phosphate pH 7.5, 1mM EDTA and 500 $\mu$ l SATA (15mg/ml dimethylsulfoxide) for 30 min at room temperature. The collagen sheets were then washed four times with phosphate buffer. Just before coupling collagen-SATA slices to APase-MHS (see below), the slices were  
10        deacetylated by incubation in a solution of 5ml 0.05M phosphate pH 7.5, 1mM EDTA and 500 $\mu$ l deacetylation solution (1.75g Hydroxylamine HCl and 0.475g EDTA in 50ml 0.05M phosphate pH 7.5) at room temperature. After 2h incubation the solution was decanted.

15        B: MHS (maleimidiohexanoly-N-hydroxysuccinimide or N-succinimidyl-6-maleidocaproate, Fluka) was coupled to APase as described by Peeters *et al.*, (Immunol. Methods 120, 133-143, 1989):

20        To 10mg APase/ml 0.05M phosphate pH 8.0 was added 1.54mg MHS/40 $\mu$ l dimethyl-formamide. After 5 min incubation at room temperature 1ml 0.05M phosphate pH 6.0 was added and the maleimidated APase separated from the smaller, organic molecules by means of a Sephadex G-25 column (10 x 1cm). The column was eluted with 0.05M phosphate pH 6.0 and fractions of 1.5ml were  
25        collected. The APase activity containing fractions in the void volume were pooled (ca 4ml).

C: Reaction between collagen-SATA slices and APase-MHS:

30        To the deacetylated collagen-SATA slices 4ml of APase-MHS in 0.05M phosphate pH 6.0 were added and the slices incubated for 4h at room temperature. Afterwards the solution was decanted and the slices washed as described in Example 1. The slices were stored in glycine buffer at 4°C. The dentinal collagen slices contained  $1.36 \pm 0.28$  mU APase/ $\mu$ g hypro, the bone collagen slices  $2.39 \pm 0.41$  mU APase/ $\mu$ g hypro.

- 17 -

RESULTS

The results of this experiment are summarised in Figure 1.

The symbols used are:

	-O-	dentin/skull	OF	older females
5	-∇-	dentin/abdomen	OM	older males
	-Δ-	bone/abdomen	YF	younger females
	-+	bone/skull	YM	younger males

The results show that mineralisation of the collagen slices (Y axis) was greatly influenced by sex, age and phosphoprotein (PP) content. Especially the bone-derived slices (low in PP content) proved to be quite low in their degree of mineralisation when compared with the dentin-derived slices (relatively high in PP content). Male rats showed more influx of mineral (as measured by phosphate content) than females. The younger animals showed more mineral uptake than the older ones. A high correlation was found between mineralisation of the implant and the concentration of inorganic phosphate in the serum (X axis).

Only minor differences were seen between the various implantation sites, the dentin-derived slices in the skull showing a slightly higher degree of mineralisation than those in the back and the abdomen. No differences were seen between the lower back region and the abdominal wall.

All control slices had remained free of mineral within the 2 week experimental period.

- 18 -

CLAIMS

1. A material useful in the repair of the skeleton which comprises a biocompatible carrier material in combination with an amount of a phosphatase enzyme which is sufficient to promote mineralisation of the material.
2. A material according to claim 1 characterised in that it exhibits a level of enzyme activity of at least 0.5 milliunits phosphatase per 1.0 microgram of hydroxyproline.
3. A material according to claim 2 characterised in that it exhibits a level of enzyme activity of at least 1.0 milliunits phosphatase per 1.0 microgram of hydroxyproline.
4. A material according to any of claims 1 to 4 characterised in that the phosphatase enzyme is an alkaline phosphatase.
5. A material according to any of claims 1 to 4 characterised in that the phosphatase enzyme is covalently bonded to the carrier.
6. A material according to claim 5 characterised in that the bonding is effected using a polyfunctional coupling agent.
7. A material according to claim 6 characterised in that the coupling agent is selected from the group comprising glutaraldehyde, 1-ethyl-3(3-dimethylaminopropyl)carbodiimide and SATA-MHS (succinimidyl-S-acetylthioacetate and maleimido-hexanoyl-N-hydroxysuccinide).
8. A material according to claim 7 characterised in that the coupling agent is SATA-MHS.
9. A material according to any of the preceding claims characterised in that it further comprises combined organic or inorganic phosphate, phosphoprotein and/or dentinal phosphophoryns.
10. A material according to claim 8 characterised in that it comprises at least 0.03 microgrammes of phosphate per microgram of hydroxyproline.
11. A material according to claim 9 characterised in that the carrier material inherently comprises at least 0.03 micrograms of phosphate per microgram of hydroxyproline.

- 19 -

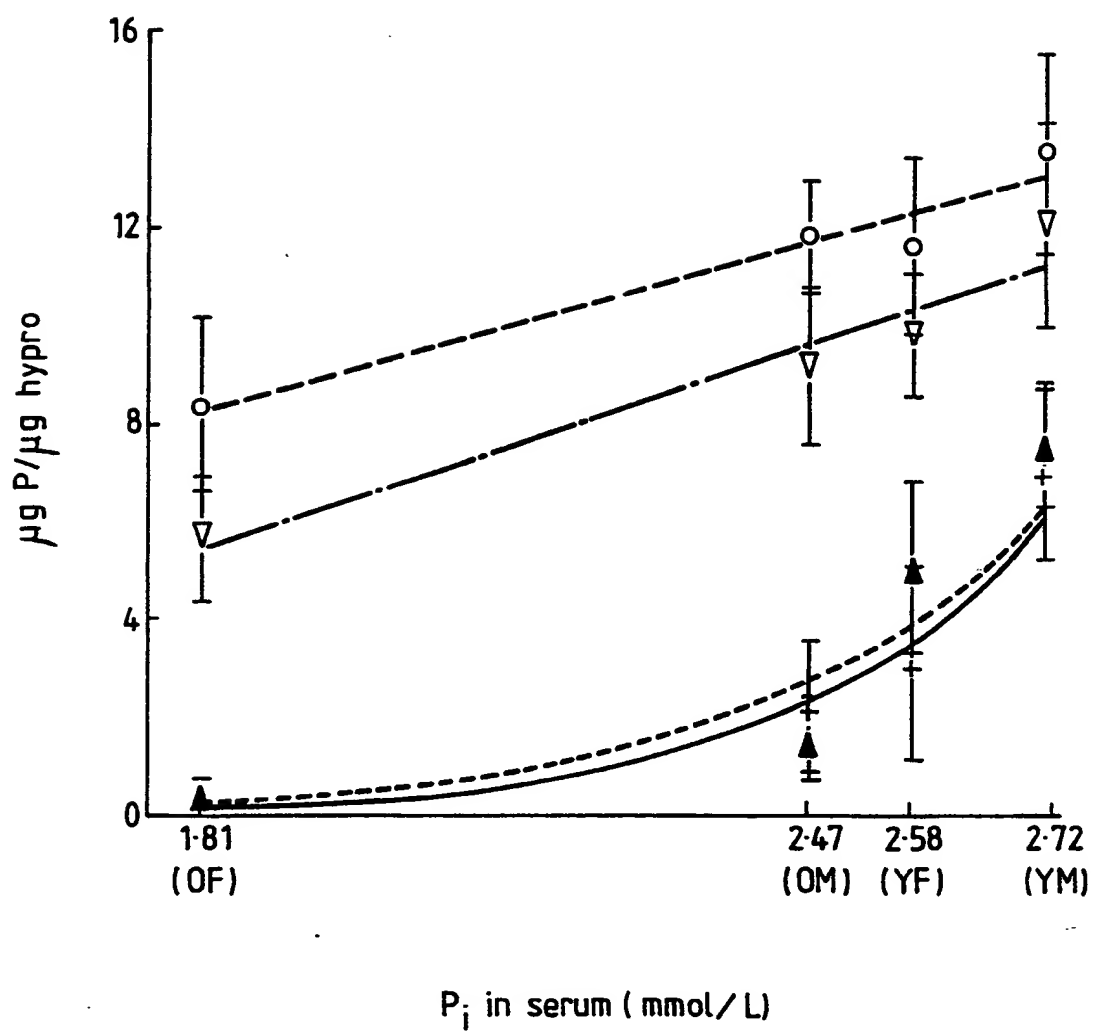
12. A material according to claim 9 characterised in that an additional quantity of organophosphate, phosphoprotein or dentinal phosphophoryns is combined with the carrier material.
13. A material according to any of claims 9 to 11 characterised  
5 in that an additional quantity of calcium ions and inorganic phosphate is combined with the carrier material.
14. A material according to any of the preceding claims characterised in that the biocompatible carrier material has been produced by the demineralisation of calcified human or  
10 animal tissue.
15. A material according to claim 14 characterised in that the carrier comprises at least a substantial proportion of fibrillar collagen.
16. A material according to any of claims 1 to 15 characterised  
15 in that the carrier comprises demineralised dentin.
17. A material according to either of claims 1 to 15 characterised in that the carrier comprises demineralised bone.
18. A material according to any of the preceding claims characterised in that it comprises at least 200 units of  
20 phosphatase per cubic centimetre.
19. A process for the production of a material according to any of claims 1 to 18 characterised in that the biocompatible carrier and the enzyme are incubated in a solution of a polyfunctional coupling agent.
- 25 20. A process according to claim 19 characterised in that the polyfunctional coupling agent is glutaraldehyde.
21. A process according to claim 19 characterised in that the polyfunctional coupling agent is 1-ethyl-3(3-dimethylaminopropyl) carbodiimide.
- 30 22. A process according to claim 19 characterised in that the biocompatible carrier is incubated in a solution of succinimidyl-S-acetylthioacetate and the enzyme is incubated with maleimidohexanoyl-N-hydroxysuccinimide and the carrier and enzyme are allowed to react.

- 20 -

23. A process according to any of claims 8 to 22 characterised in that the incubation is carried out in the presence of additional organophosphate compounds.
24. A process according to claim 22 characterised in that the organophosphate compounds are phosphoproteins or dentinal phosphophoryns.
25. A process according to any of claims 8 to 23 characterised in that the incubation is carried out in the presence of calcium and inorganic phosphate ions.
26. A method for the treatment of the skeleton of an animal in order to promote mineralisation and the growth of bone which comprises the insertion of an implant comprising a material according to any of claims 1 to 17.
27. A method according to claim 26 characterised in that the concentration of organophosphorus compounds in the locality of the implant is supplemented from a source outside the body.

1 / 1

FIG. 1



# INTERNATIONAL SEARCH REPORT

International Application No

PCT/GB 92/01247

## I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all)<sup>6</sup>

According to International Patent Classification (IPC) or to both National Classification and IPC  
 Int.C1.5                      A 61 L 27/00

## II. FIELDS SEARCHED

### Minimum Documentation Searched<sup>7</sup>

Classification System	Classification Symbols
Int.C1.5	A 61 L

Documentation Searched other than Minimum Documentation  
 to the Extent that such Documents are Included in the Fields Searched<sup>8</sup>

## III. DOCUMENTS CONSIDERED TO BE RELEVANT<sup>9</sup>

Category <sup>10</sup>	Citation of Document, <sup>11</sup> with indication, where appropriate, of the relevant passages <sup>12</sup>	Relevant to Claim No. <sup>13</sup>
X	US,A,4409332 (S.R. JEFFERIES et al.) 11 October 1983, see column 2, lines 16-20,43-68; column 3, lines 6-26 (cited in the application) ---	1-27
X	US,A,4394370 (S.R. JEFFERIES) 19 July 1983, see column 2, lines 32-62; example III (cited in the application) ---	1-27
X	Journal of Dental Research, vol. 70, no. 3, March 1991, W. BEERTSEN et al.: "Alkaline phosphatase induces the deposition of calcified layers in relation to dentin: an in vitro study to mimic the formation of afibrillar acellular cementum", pages 176-181, see discussion --- -/-	1-27

### <sup>10</sup> Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"A" document member of the same patent family

## IV. CERTIFICATION

Date of the Actual Completion of the International Search

26-08-1992

Date of Mailing of this International Search Report

30. 09. 92

International Searching Authority

EUROPEAN PATENT OFFICE

Signature of Authorized Officer

*Dagmar Frank*



III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)		
Category *	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No.
A	Matrix, vol. 9, no. 2, March 1989, Gustav Fischer Verlag, (Stuttgart, DE), W. BEERTSEN et al.: "Calcification of dentinal collagen by cultured rabbit periosteum: the role of alkaline phosphatase", pages 159-171, see summary (cited in the application) ----	1-27
A	GB,A,2055848 (E.N.I.) 11 March 1981, see claims ----	1-27
A	Chemical Abstracts, vol. 97, no. 10, 6 September 1982, (Columbus, Ohio, US), see page 425, abstract no. 78947m, & JP,A,8289867 (K. MIURA) 4 June 1982, see abstract ----	1
A	NL,A,8902155 (STICHTING BIOMAT TE GRONINGEN) 18 March 1991 -----	

**Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)**

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.:  
because they relate to subject matter not required to be searched by this Authority, namely:  
**ALTHOUGH CLAIMS 26-27 ARE DIRECTED TO A METHOD OF TREATMENT OF THE HUMAN/  
ANIMAL BODY THE SEARCH HAS BEEN CARRIED OUT AND BASED ON THE ALLEGED  
EFFECTS OF THE COMPOUNDS.**
2. ☐ Claims Nos.:  
because they relate to parts of the international application that do not comply with the prescribed requirements to such  
an extent that no meaningful international search can be carried out, specifically:
3. ☐ Claims Nos.:  
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

**Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)**

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all  
searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment  
of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report  
covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is  
restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

**ANNEX TO THE INTERNATIONAL SEARCH REPORT  
ON INTERNATIONAL PATENT APPLICATION NO.**

GB 9201247  
SA 61978

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report. The members are as contained in the European Patent Office EDP file on 24/09/92. The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
US-A- 4409332	11-10-83	CA-A- 1186625	07-05-85
US-A- 4394370	19-07-83	US-A- 4472840	25-09-84
GB-A- 2055848	11-03-81	AT-B- 387906	10-04-89
		AU-B- 536483	10-05-84
		AU-A- 6004980	05-02-81
		BE-A- 884301	14-01-81
		CA-A- 1148466	21-06-83
		CH-A- 645920	31-10-84
		DE-A, C 3026805	05-02-81
		FR-A, B 2462475	13-02-81
		JP-B- 1010228	21-02-89
		JP-C- 1528489	30-10-89
		JP-A- 56072868	17-06-81
		LU-A- 82623	02-02-81
		NL-A- 8004068	03-02-81
		SE-A- 8005061	02-02-81
		US-A- 4396716	02-08-83
NL-A- 8902155	18-03-91	None	